



WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

(51) International Patent Classification: Not classified		A2	(11) International Publication Number: WO 90/11676
			(43) International Publication Date: 18 October 1990 (18.10.90)
(21) International Application Number: PCT/US90/01864 (22) International Filing Date: 5 April 1990 (05.04.90) (30) Priority data: 334,459 6 April 1989 (06.04.89) US (71) Applicant: CENTOCOR, INC. [US/US]; 244 Great Valley Parkway, Malvern, PA 19355 (US). (72) Inventors: DADDONA, Peter, E. ; 642 Marydell Drive, West Chester, PA 19380 (US). BERGER, Harvey, J. ; 687 Wetherby Lane, Devon, PA 19333 (US). McEVER, Rodger, P. ; 1716 Guilford Lane, Oklahoma City, OK 73120 (US).		(74) Agents: DECONTI, Giulio, A., Jr. et al.; Hamilton, Brook, Smith & Reynolds, Two Militia Drive, Lexington, MA 02173 (US). (81) Designated States: AT (European patent), BE (European patent), CA, CH (European patent), DE (European patent), DK (European patent), ES (European patent), FR (European patent), GB (European patent), IT (European patent), JP, LU (European patent), NL (European patent), SE (European patent). Published With declaration under Article 17(2)(a). Without classification and without abstract; title not checked by the International Searching Authority.	
(54) Title: METHOD FOR IMAGING ATHEROSCLEROTIC PLAQUE			
ATTORNEY DOCKET NUMBER: 10177-211-999 (Cam.no. 008563-999208 SERIAL NUMBER: 00/010 288			

ATTORNEY DOCKET NUMBER: 10177-211-999
(Cam no: 008563-999208
SERIAL NUMBER: 09/910,388
REFERENCE: B24

BEST AVAILABLE COPY

DESIGNATIONS OF "DE"

Until further notice, any designation of "DE" in any international application whose international filing date is prior to October 3, 1990, shall have effect in the territory of the Federal Republic of Germany with the exception of the territory of the former German Democratic Republic.

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	ES	Spain	MG	Madagascar
AU	Australia	FI	Finland	ML	Mali
BB	Barbados	FR	France	MR	Mauritania
BE	Belgium	GA	Gabon	MW	Malawi
BF	Burkina Faso	GB	United Kingdom	NL	Netherlands
BG	Bulgaria	HU	Hungary	NO	Norway
BJ	Benin	IT	Italy	RO	Romania
BR	Brazil	JP	Japan	SD	Sudan
CA	Canada	KP	Democratic People's Republic of Korea	SE	Sweden
CF	Central African Republic	KR	Republic of Korea	SN	Senegal
CG	Congo	LJ	Liechtenstein	SU	Soviet Union
CH	Switzerland	LK	Sri Lanka	TD	Chad
CM	Cameroon	LU	Luxembourg	TG	Togo
DE	Germany, Federal Republic of	MC	Monaco	US	United States of America
DK	Denmark				

-1-

METHODS FOR IMAGING ATHEROSCLEROTIC PLAQUEDescriptionBackground of the Invention

05 A generally accepted theory of development of
atherosclerotic plaque is based on injury to the
endothelial cells which line the inside surface of a
blood vessel. The initial injury can be caused by
chemical factors related to chronically elevated
10 levels of cholesterol in the blood or by mechanical
stress associated with hypertension.

The clinical effects of atherosclerotic plaque
are due to its space occupying characteristics which
lead to stenosis (narrowing) or its thrombogenic
15 (clot-causing) characteristics which appear related
to rupture of the fibrous cap and the resulting
ulceration of the plaque surface. As a result, a
thrombus will form at the site of the plaque. This,
in turn, will almost invariably lead to acute
20 myocardial infarction if present in coronary
arteries. See Friedman et al., Am. J. Pathol.,
48:19-25 (1966); Schwartz, C.J. Atherosclerosis,
15:1-4 (1972).

It appears that the first stage of plaque
25 formation is caused by macrophages migrating through
the injured endothelium and accumulating subendo-
thelially. The lesion is enlarged by an accumula-
tion of smooth muscle cells which combine with the
macrophages to form foam cells in fatty streaks.
30 Endothelial cells appear to then separate, exposing

-2-

much of the fatty streak to the circulation. This ulcerated condition provides opportunities for platelet aggregation and mural thrombosis. Damage to the endothelium and the resulting platelet
05 activation may also cause further enlargement of the plaque.

The important consequence of these effects is reduction of blood flow to the affected organ (heart, brain, etc.). Atherosclerosis is the
10 underlying cause of 75% of the one million deaths from cardiovascular disease in the United States each year. Atherosclerosis accounts for a large proportion of heart attacks, many strokes, most aneurisms of the lower abdominal aorta, and many
15 cases of peripheral vascular disease. Myocardial infarcts have been imaged with antibodies that localize at the infarct site. See, e.g., Haber et al., U.S. Patent 4,036,945. A need exists for imaging methods of high reliability and high
20 resolution that can be used for diagnosing and monitoring atherosclerotic lesions. This is especially true for those lesions that have ulcerated or fissured, resulting in formation of blood clots.

25 Summary of the Invention

This invention pertains to an method of imaging atherosclerotic plaques in vivo. The method entails the use of monoclonal antibodies specific for activated platelets or activated endothelial
30 cells. According to the method of this invention, a

- 3 -

radiolabeled monoclonal antibody or antibody
fragment specific for activated platelets, such as
the monoclonal antibody S12 which is specific for
the GMP-140 glycoprotein of activated platelets or
05 activated endothelial cells, is administered to a
patient suspected of having an endothelial injury
such as an ulcerated atherosclerotic lesion. The
radiolabeled monoclonal antibody is allowed to
accumulate at the site of the plaque. Thereafter,
10 the signal generated by the radiolabel is detected
by a photoscanning device and the signal is
converted to an image of the atherosclerotic plaque.

Brief Description of Figures

Figure 1 shows schematic representations of
15 planar imaging of technetium-99m (Tc-99m) S12 Fab'
in rabbits. The increasing intensity of radiolabel
uptake is shown by the direction of the arrow in the
figure legend.

Figure 1A shows a schematic diagram of Tc-99m
20 S12 Fab' uptake and distribution in a control
rabbit.

Figure 1B shows a schematic diagram of Tc-99m
S12 Fab' uptake and distribution in an
25 atherosclerotic rabbit with repeat endothelial
injury.

Figure 2 shows a schematic diagram of Tc-99m
S12 Fab' uptake and localization in the aorta of an
atherosclerotic rabbit.

Figure 3 shows a schematic representation of
30 Tc-99m S12 Fab' uptake and distribution in the

-4-

excised aorta of the same rabbit as shown in the previous Figure.

Detailed Description of the Invention

An immunoscintigraphic image of an
05 atherosclerotic plaque can be obtained with
radiolabeled antibodies specific for activated
platelets or activated endothelial cells. An
antibody or antibody fragment (antigen binding
fragment) specific for activated platelets or
10 activated endothelial cells conjugated to a label
which generates a signal detectable by external
scintigraphy is administered to a patient suspected
of having an atherosclerotic plaque. Sufficient
time is allowed for the labeled antibody to localize
15 at the site or sites of plaque. The signal
generated by the label is detected by a
photoscanning device and the signal is then
converted to an image of the atherosclerotic plaque.

Preferred antibodies for plaque imaging are
20 specific for an epitope of the GMP-140 protein, a
marker of activated platelets. The protein is
localized in alpha-granule membranes in unstimulated
platelets. After thrombin stimulation and membrane
fusion, the protein is redistributed to the cell
25 surface and plasma membrane. The protein, GMP-140
has been described by Stenberg et al., J. Cell
Biol., 101, 880 (1985).

In addition to being highly specific for
activated platelets, antibodies specific for the
30 GMP-140 glycoprotein have another advantage. The

- 5 -

antibodies also react with endothelial GMP-140. Endothelial GMP-140, like its platelet counterpart, is localized to the membranes of secretory storage granules in unstimulated endothelial cells. When
05 these cells are activated, the protein redistributes to the cell surface, just as it does in activated platelets. Thus, GMP-140 can serve a role as a marker of activated endothelial cells damaged by atherosclerotic ulceration or other endothelial
10 injury. Imaging methods of this invention employing platelet-specific monoclonal antibodies are able to detect sites of endothelial injury in which vascular lesions are associated with platelet activation. These ulcerated or ruptured sites of plaque
15 formation can be detected using platelet-specific monoclonal antibodies.

A particularly preferred antibody for the method of this invention is the monoclonal antibody S12 which reacts specifically with the GMP-140
20 protein. The S12 antibody reacts minimally with unstimulated human platelets but binds extensively after platelets have been activated with thrombin. See McEver and Martin, J. Biol. Chem., 259:9799 (1984) incorporated herein by reference.

25 Antibodies against components of activated platelets other than GMP-140 may also be used. For example, the monoclonal antibody Tab, which binds to platelet glycoprotein IIb, can also be used in embodiments of this invention. See McEver et al.,
30 J. Biol. Chem., 258:5269 (1983), incorporated herein by reference.

-6-

GMP-140 specific monoclonal antibodies of this invention are produced by hybrid cell lines commonly known as hybridomas. These hybrid cells are formed by fusion of an anti-GMP-140 antibody-producing cell and an immortalizing partner (i.e. a cell line which imparts long term tissue culture stability to the hybrid cell). In the formation of the hybrid cell lines, the anti-GMP-140 antibody-producing cell can be a B lymphocyte obtained from an animal or human who has developed an immune response to GMP-140: (i) naturally; (ii) as a result of a pathologic process; (iii) as a consequence of having been immunized with material containing GMP-140 such as activated platelets, activated endothelial cells, or a biological preparation comprising GMP-140. The immortalizing partner can be a cell of B lymphocyte lineage such as a B lymphoblastoid cell line or a plasmacytoma cell such as a myeloma cell, itself a antibody producing cell that is also malignant.

In particular, murine hybridomas producing GMP-140-specific monoclonal antibodies are formed by fusion of: (i) mouse myeloma cells and (ii) spleen cells from mice immunized against GMP-140 positive platelets or endothelial cells, purified GMP-140, or other biological preparations containing GMP-140. Fusions are accomplished by standard procedures well known in the art. Kohler and Milstein, Nature, 256: 495-497 (1975) and Kennet, Monoclonal Antibodies (Kennet et. al., eds.), pp. 365-367, Plenum Press, N.Y. 1980). The resulting murine hybridoma clones are then screened for production of antibody

-7-

reactive with GMP-140-positive platelets, endothelial cells, or preparations containing GMP-140. Those which secrete antibodies of the appropriate reactivity and specificity are cloned to
05 yield a homogeneous cell line secreting anti-GMP-140 antibody.

Human hybridomas which produce monoclonal anti-GMP-140 antibodies are formed from the fusion of: (i) human B cells from an individual producing
10 anti-GMP-140 antibodies and (ii) human B lymphoblastoid or human/human-mouse myeloma cells. The fusion partner for the myeloma cell may be an anti-GMP-140-producing B lymphocyte derived from the blood, spleen, or tonsil lymph node. The fusion and
15 screening techniques are essentially the same as those used in the production and screening of murine anti-GMP-140 generating hybridomas.

Mouse/human hybridomas which produce human anti-GMP-140 antibody can be formed from the fusion
20 of human antibody-producing cells and a murine plasmacytoma cell or a cell which itself is a hybrid having the appropriate properties such as the ability to fuse with human lymphocytes at a high frequency; support the synthesis and secretion of
25 high levels of antibody; and support secretion of antibody for prolonged periods of time in culture.

Another way of forming the anti-GMP-140-producing cell line is by transformation of antibody-producing cells. For
30 example, an anti-GMP-140 producing B lymphocyte may be infected and transformed with a virus such as

- 8 -

Epstein-Barr virus to yield an immortal anti-GMP-140 producing cell. Kozbor and Roder, Immunology Today, 4, 72-79 (1983). Alternatively, the B lymphocyte may be transformed by a transforming gene or gene product.

GMP-140-specific monoclonal antibodies are produced in large quantities by injecting anti-GMP-140 antibody-producing hybridomas into the peritoneal cavity of mice or other appropriate hosts and, after appropriate time, harvesting the resulting ascitic fluid which contains a high titer of antibody, and isolating the desired anti-GMP-140 monoclonal antibody therefrom. Allogeneic or xenogeneic hybridomas can be injected into immunosuppressed, irradiated or athymic nude mice. Alternatively, the antibodies may be produced by culturing anti-GMP-140 producing cells in vitro and isolating secreted monoclonal anti-GMP-140 antibodies from the cell culture medium.

Antibody fragments, rather than whole antibody molecules, are generally preferred for use in the plaque-imaging method of this invention. Because they are distributed more readily in the tissues than are entire antibody molecules, antibody fragments accumulate at the target site more rapidly. Thus an image can be obtained in less time than is possible using whole antibody. These fragments are also cleared more rapidly from the circulation, resulting in a lower background signal. Haber et al., U.S. Patent No. 4,036,945; Goldenberg et al., U.S. Patent No. 4,331,647. The antigen

- 9 -

binding fragments F_v , Fab, Fab' and $F(ab')_2$ or
analogs of these fragments can be used. The Fab
fragment can be prepared by digestion of whole
immunoglobulin molecules with the enzyme papain; the
05 $F(ab')_2$ fragment can be prepared by digestion of the
whole immunoglobulin molecule with pepsin; and the
Fab' fragment produced from the $F(ab')_2$ fragment by
reduction with dithiothreitol or cysteine, according
to any of several well known protocols. In
10 addition, fragments can also be prepared by
recombinant DNA techniques. See, for example, U.S.
Patent Application Serial No. 195,720 (Coller and
Knight), filed May 18, 1988, the teachings of which
are incorporated by reference herein.

15 The antibodies or antibody fragments can be
labeled with radioisotopes such as Indium-111 and
Technetium-99m which are suitable for detection by
external scintigraphy. The gamma-emitters
Indium-111 and Technetium-99m are preferred because
20 these radiometals are detectable with a gamma camera
and have favorable half-lives in vivo. Technetium-
99m is an ideal radionuclide for scintigraphic
imaging because of its nuclear properties. It has a
single photon energy of 140 KeV, a half-life of
25 about 6 hours, and it is readily available from a
 ^{99}Mo - ^{99m}Tc generator.

Antibodies can be labeled by any of the many
techniques known in the art. The preferred
technique for technetium-99m labelling is that of
30 Pak, K.Y et al., United States Patent Application
Serial No. 034,003, filed April 2, 1987, the

-10-

teachings of which are incorporated herein.
Antibody can also be labeled with a radiometal via a
chelating agent such as diethylene-

05 triaminepentaacetic acid (DTPA) which is conjugated
to the antibody. In this indirect labeling
approach, the radiometal is chelated by the
chelating agent attached to the antibody. See, for
example, Khaw et al., Science, 209:295-297 (1980)
Krejcarek and Tucker, Biochem. Biophys. Res.
10 Commun., 77:581-585 (1977); Childs, and Hnatowich,
J. Nucl. Med., 26:293 (1985).

Various subtraction techniques can be used to
enhance detection and/or localization of labeled
antibodies. See, for example, Ballou, B. et al.,
15 Science, 206:844-847 (1979), and references cited
therein.

The labeled antibody is administered to a
patient in the form of an injectable composition.
Injection may be intravenous or intraarterial. A
20 typical composition contains a quantity of
radiolabeled antibody sufficient for imaging (e.g.,
specific activity 3-150 mCi/mg) in neutral phosphate
buffer containing physiological saline.

Photoscanning can be performed by standard
25 techniques. Goldenberg et al., New Engl. J. Med.,
298:1389 (1978) and references cited therein.

The method of this invention can be used to
distinguish between ulcerated and nonulcerated
atherosclerotic plaque. The method is clinically
30 useful for monitoring the extent of progression of
atherosclerosis in heart attack survivors and in

-11-

screening a population at risk for atherosclerosis (about 143 million worldwide). Information provided by the methods of this invention will aid physicians in evaluation of therapy and in early diagnosis of atherosclerotic plaque.

This invention will now be more fully disclosed according to the following examples.

Example I: Preparation of Technetium 99m-labeled anti-platelet Fab' fragment

10 A. Preparation of S12 Fab'

S12 IgG is purified from hybridoma tissue culture supernatant using Protein A Sepharose column chromatography. The F(ab')₂ fragment is prepared from the IgG by pepsin proteolytic digestion at pH 3.9 and subsequently purified by cation exchange chromatography using for example, Pharmacia Fast Flow S. The Fab' fragment is produced from the F(ab')₂ by reduction with dithiothreitol, purified by diafiltration or column gel filtration and formulated at 1-2 mg/ml in 50mM sodium phosphate buffer pH 6.3 containing 100mM sodium chloride and 1mM EDTA.

B. Labeling of S12 Fab' with Tc-99m

The S12 Fab' antibody fragment is labeled with Technetium 99-m as described by Pak et al., Circulation 76 Supplement II, #2013 (1987); Pak et al U.S. Patent Application Serial No. 034,003 (filed April 2, 1987). Using D-glucarate as the transfer ligand, 200-400 µg aliquots of the Fab' are labeled

-12-

with Tc-99m to a specific activity of 10-40 mCi/mg. Final protein concentrations are 0.5 - 1 mg/ml in the labeling solution. The quantitative incorporation of Tc-99m is determined by gel
05 filtration HPLC equipped with a radiometric detector, ITLC and paper chromatography.

Immunoactivity of Tc-99m S12 Fab' is evaluated using a platelet binding assay with thrombin-activated or unactivated platelets.

10 In this method, platelets are isolated by differential centrifugation from ACD treated whole human blood taken from normal volunteers. In the assay a fixed number of washed platelets (5×10^7 /ml) in PBS with 1% bovine serum albumin (BSA) is
15 incubated with varying concentrations of Tc-99m S12 Fab' or Tc-99m labeled control antibody in PBS with BSA, 1 mM EDTA (final conc.) to prevent platelet aggregation and 8 units/ml of bovine thrombin (final conc.). Thrombin is omitted from the assay when
20 binding to resting platelets is evaluated. Incubations are performed at room temperature for 60-120 min. Platelets are washed by centrifugation and radioactivity bound to platelets determined using a gamma counter set for Tc-99m.

25 Specific saturable binding of Tc-99m S12 Fab' has been demonstrated using this assay. The specificity of binding is such that 200 ug/mL of unlabeled S12 IgG prevents the binding of Tc-99m S12 Fab' to attached platelets. The binding is also
30 activation dependent since there is only a background level of binding in this assay using

-13-

platelets which have not been exposed to thrombin. The labeled S12 Fab' reacts strongly with activated platelets isolated from human, baboon, and rabbit. The antibody shows weak to no reactivity with
05 unactivated platelets from these same species.

Example II: Imaging using Tc-99m S12 Fab' monoclonal antibody

A. Preparation of a Rabbit Atherosclerosis Model

Rabbits weighing 5-10 lbs. received a 2%
10 cholesterol atherogenic diet in order to elevate serum cholesterol levels. The diet consisted of rabbit chow supplemented by cholesterol-enriched peanut oil to produce hypercholesterolemia (cholesterol - 500-1000 mg %). This control group
15 was kept on the diet for 1-2 weeks. A more severe atherosclerotic condition was induced in the test group using the Baumgartner technique. Baumgartner H.R., Ges. Exp. Med. 137, 227, (1963). In brief, the endothelial lining of the infra-renal aorta is
20 stripped or denuded by percutaneous placement of a 4 French Fogarty catheter via a femoral arteriotomy under sterile conditions. The catheter is retrogradely advanced through the iliac artery under
25 fluoroscopic guidance. The balloon tip is inflated with saline and then drawn three times from the level of the renal to the proximal right iliac artery. After endothelial injury, the high cholesterol atherogenic diet is continued for an
30 additional 4-6 weeks until the arterial re-injury phase of the study. This model has previously been

-14-

shown to cause significant atherosclerosis characterized by intimal thickening, foam cell lesions and neovascularization in 66% of the animals. Baumgartner H.R., Ges. Exp. Med. 137,227(1963).

B. Tc-99m Fab' Imaging of Arterial Injury in Rabbit Atherosclerosis Model

All rabbits were modeled to have atherosclerosis of the lower aorta and proximal iliac artery using a Fogarty catheter injury followed by high cholesterol diet for 4-6 weeks, as described previously. Rabbits were divided into three groups of increasingly severe aortic injury as follows:

- 15 I. Control group (resting plaque);
- II. Reinjury group (repeated balloon injury causing focal ruptured plaque and endothelial damage); and
- 20 III. Percutaneous transluminal angioplasty (PTA) with intravascular stent placement (repeated balloon injury and placement of a 1cm long wire mesh stent).

At the time of each imaging study, contrast angiography was performed to confirm the site of injury. Animals were not anticoagulated or pre-treated with antiplatelet medications.

-15-

Within 15 minutes of injury, each rabbit was injected with 5 mCi/400 μ g of Tc-99m S12 Fab' via an ear vein. Serial 5 minute images were obtained for 60 minutes after injection using a Searle 37 x gamma camera and high resolution collimator and analyzed on a MDS A2 computer. In vivo Tc-99m S12 Fab' uptake was quantitated 45 - 60 minutes post injection from planar image regions-of-interest to derive an injured to normal (I:N) aortic activity ratio. Results from the three subgroups of increasingly severe injury are shown in Table 1.

-16-

Table 1

	<u>Group</u>	<u>Rabbits</u>	<u>Injury</u>	<u>S12 I:N Ratio</u>
	I.	4	Control	1.2 \pm 0.17
	II.	6	Reinjury	1.5 \pm 0.18
05	III.	3	PTA/Stent	1.9 \pm 0.12

The reinjury (II) and PTA/Stent (III) groups showed distinct focal Tc-99m S12 Fab' uptake that was significantly greater than that of control group I ($p < 0.01$). Ex vivo imaging of the blood-free
10 excised aortas confirmed marked S12 localization with an I:N = 3.2 ± 1.4 at pathologically and angiographically proven injury sites.

These results demonstrate that Tc-99m S12 Fab' localizes sites of active platelet degranulation in
15 balloon-injured atherosclerotic endothelium with high specificity. Further, the degree of S12 uptake is proportional to the severity of the arterial injury.

Intense and focal uptake of Tc-99m S12 Fab' was
20 demonstrated in the reinjury and PTA stented atherosclerotic model relative to the control atherosclerotic rabbits using planar imaging. Fig. 1 shows a schematic diagram of the planar imaging of Tc-99m S12 Fab' in a representative control rabbit
25 (Figure 1A) and an atherosclerotic rabbit with repeat Fogarty injury (Figure 1B). The control rabbit was imaged 35 min. after Tc-99m S12 Fab'

-17-

injection and shows non-specific radiochemical uptake in the two kidneys (upper quadrant), the spleen (left mid-section) and the bladder (lower center). Minimal uptake is visible in the descended
05 infra renal aorta (mid-line area). An atherosclerotic rabbit with Fogarty treatment (Figure 1B) shows intense radiochemical uptake in the upper infra renal aorta between the two kidneys (upper quadrant). Angiographic analysis of these
10 rabbits (not presented here) clearly reveal the bulging aorta of the atherosclerotic rabbit relative to the control.

A complete set of in vivo and ex vivo images, angiograms, x-rays and pathology pictures was taken
15 from a rabbit that received a PTA stent following atherosclerotic modeling and previous Fogarty balloon injury. Figure 2 shows a schematic diagram of the radiolocalization of the Tc-99m S12 Fab' in the descending aorta of this rabbit taken 35 minutes
20 after injection. Localization is noted as the intense streak in the area of the descending aorta. Sixty minutes after imaging, the rabbit was sacrificed, the descending aorta was excised, washed free of blood and mounted for pathological
25 examination. Visual analysis revealed a mixture of white and red thrombus in the wire mesh of the stent and extensive atherosclerosis over the entire region of the aorta. A representation of the ex vivo image of the excised aorta indicating the localization of
30 the Tc-99m S12 Fab' is shown in Figure 3. These data, taken together show an excellent correlation

-18-

of the atherosclerotic injury site by angiography, pathology and in vivo and ex vivo imaging using the Tc-99m S12 Fab' antibody.

Equivalents

- 05 Those skilled in the art will recognize or be able to ascertain, using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed in the
- 10 following claims.

-19-

CLAIMS

1. A method of imaging an atherosclerotic plaque,
comprising the steps of:
 - 05 a. administering to an individual suspected
of having atherosclerotic plaque, a
radiolabeled monoclonal antibody or
fragment thereof, specific for activated
platelets or activated endothelial cells;
 - 10 b. allowing the radiolabeled monoclonal
antibody or antibody-fragment to accumu-
late at the plaque site;
 - c. detecting the signal generated by the
radiolabel by means of a photoscanning
device;
 - 15 d. converting the detected signal into an
image of the plaque.
2. A method of Claim 1, wherein the monoclonal
antibody fragment is selected from the group
consisting of Fab, Fab', F(ab')₂, and F_v
20 fragments.
3. A method of Claim 1, wherein the monoclonal
antibody is labeled with a radioactive material
selected from the group consisting of
Technetium-99m and Indium-111.

-20-

4. A method of Claim 1, wherein the activated platelet-specific monoclonal antibody is specific for the activated platelet membrane protein GMP-140.
- 05 5. A method of Claim 4, wherein the monoclonal antibody is S12.
6. A method of claim 1, wherein the atherosclerotic plaque is ulcerated or ruptured.
- 10 7. A method of imaging atherosclerotic plaque, comprising the steps of:
 - a. administering to an individual suspected of having atherosclerotic plaque, a ^{99m}Tc-labeled Fab fragment of monoclonal
15 antibody S12;
 - b. allowing the radiolabeled antibody fragment to accumulate at the plaque site;
 - c. detecting the signal generated by means of a gamma camera;
 - 20 d. converting the detected signal into an image of the plaque.

1/1

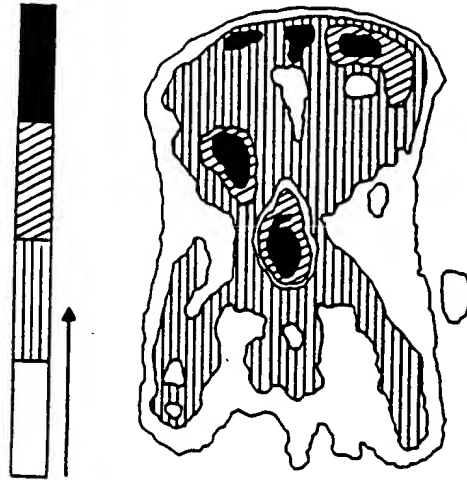


FIG. 1A

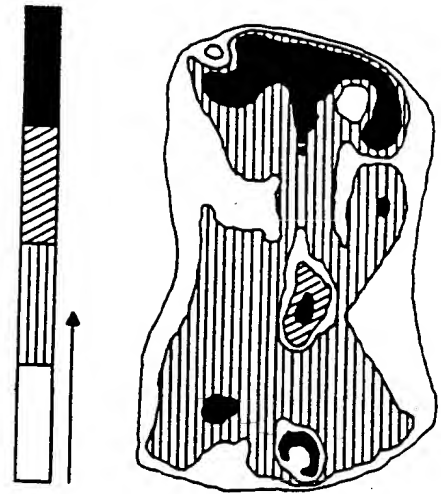


FIG. 1B

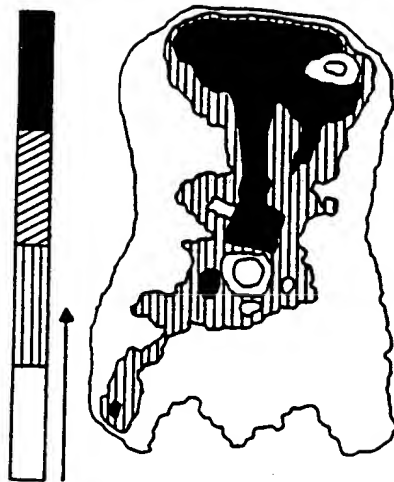


FIG. 2

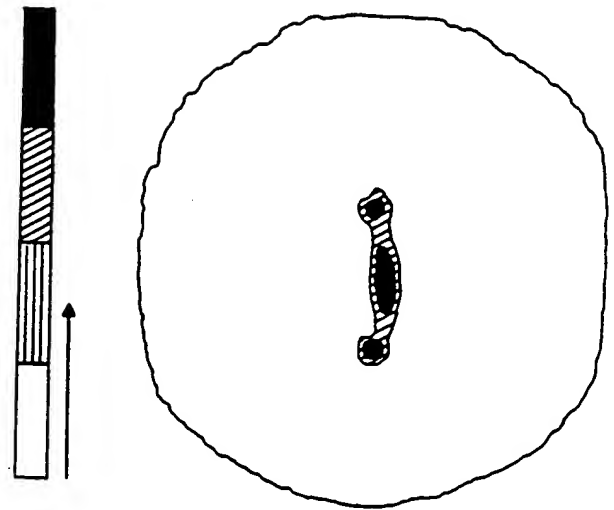


FIG. 3

PATENT COOPERATION TREATY

DECLARATION OF NON-ESTABLISHMENT OF INTERNATIONAL SEARCH REPORT

issued pursuant to PCT Article 17(2)(a) ⁽¹⁾

IDENTIFICATION OF THE INTERNATIONAL APPLICATION	APPLICANT'S OR AGENT'S FILE REFERENCE CTR88-22
International Application No. PCT/US 90/01864	International Filing Date 5th April 1990
Receiving Office RO/US	Priority Date Claimed 6th April 1989
Applicant (Name) CENTROCOR INC.	
DECLARATION	
<p>This International Searching Authority hereby declares that no international search report will be established on the above-identified international application for the reasons indicated below. (1)</p> <p>1. The subject matter of the international application relates to: (2)</p> <ul style="list-style-type: none"> a. <input type="checkbox"/> scientific theories. b. <input type="checkbox"/> mathematical theories. c. <input type="checkbox"/> plant varieties. d. <input type="checkbox"/> animal varieties. e. <input type="checkbox"/> essentially biological processes for the production of plants and animals, other than microbiological processes and the products of such processes. f. <input type="checkbox"/> schemes, rules or methods of doing business. g. <input type="checkbox"/> schemes, rules or methods of performing purely mental acts. h. <input type="checkbox"/> schemes, rules or methods of playing games. i. <input checked="" type="checkbox"/> methods for treatment of the human body by surgery or therapy. j. <input type="checkbox"/> methods for treatment of the animal body by surgery or therapy. k. <input type="checkbox"/> diagnostic methods. l. <input type="checkbox"/> mere presentations of information. m. <input type="checkbox"/> computer programs for which this International Searching Authority is not equipped to search prior art. <p>2. The failure of the following parts of the international application to comply with prescribed requirements prevents a meaningful search from being carried out: (3)</p> <ul style="list-style-type: none"> a. <input type="checkbox"/> the description. b. <input checked="" type="checkbox"/> the claims. c. <input type="checkbox"/> the drawings. <p style="margin-left: 20px;">Pls. see Rule 39.1 (iv) - PCT</p> <p>comment:</p>	
CERTIFICATION	
International Searching Authority ISA / EP	Date of Mailing - 5 -
Authorized Officer <div style="display: flex; align-items: center;"> <div style="border: 1px solid black; padding: 2px 5px; margin-right: 10px;">M. FERG</div> <div style="font-family: cursive; font-size: 1.2em;">M. Ferg</div> </div>	

**This Page is Inserted by IFW Indexing and Scanning
Operations and is not part of the Official Record**

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images include but are not limited to the items checked:

- ☐ **BLACK BORDERS**
- ☐ **IMAGE CUT OFF AT TOP, BOTTOM OR SIDES**
- ☒ **FADED TEXT OR DRAWING**
- ☐ **BLURRED OR ILLEGIBLE TEXT OR DRAWING**
- ☐ **SKEWED/SLANTED IMAGES**
- ☐ **COLOR OR BLACK AND WHITE PHOTOGRAPHS**
- ☐ **GRAY SCALE DOCUMENTS**
- ☒ **LINES OR MARKS ON ORIGINAL DOCUMENT**
- ☐ **REFERENCE(S) OR EXHIBIT(S) SUBMITTED ARE POOR QUALITY**
- ☐ **OTHER:** _____

IMAGES ARE BEST AVAILABLE COPY.

As rescanning these documents will not correct the image problems checked, please do not report these problems to the IFW Image Problem Mailbox.